

P^1, P^2 -Diimidazolyl derivatives of pyrophosphate and bis-phosphonates – synthesis, properties, and use in preparation of dinucleoside tetraphosphates and analogs†

Ivan B. Yanachkov,* Edward J. Dix, Milka I. Yanachkova and George E. Wright

Received 5th August 2010, Accepted 29th September 2010

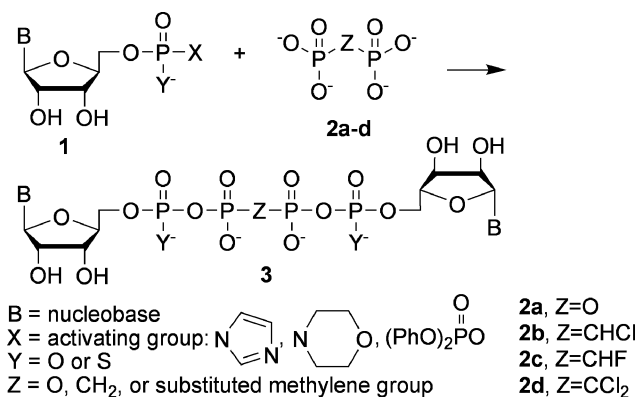
DOI: 10.1039/c0ob00542h

P^1, P^2 -Diimidazolyl derivatives of pyrophosphate and halomethylene-bis-phosphonates have been synthesized and characterized, and the mechanism of their formation was studied. These reagents enable synthesis of dinucleoside tetraphosphates and tetraphosphonates conveniently and in high yields.

Introduction

Nucleoside polyphosphates play fundamental roles in living systems, and much effort has been focused in the development of their synthetic chemistry.^{1,2} The subclass of bis-nucleoside polyphosphates, initially thought of as mere artifacts or by-products in the synthesis of mononucleoside polyphosphates, has gained increased attention in recent years,³ as they have emerged as important intra- and extracellular signal messengers which take part in the regulation of many biological processes.^{4,5} For instance, they have been implicated in regulation of blood pressure,⁶ cellular stress response,⁷ insulin and glucose levels,⁸ platelet activation,⁹ and neurotransmission.¹⁰ The class has been subject to drug development efforts,^{11–14} which resulted in late-stage clinical trials of bis-uridine tetraphosphate (Up₄U) and its analogs for treatment of cystic fibrosis¹⁵ and dry eye;¹⁶ and bis-adenosine tetraphosphate (Ap₄A) for control of blood pressure during anesthesia.¹⁷

Bis-nucleoside tetraphosphates and their phosphonate analogs (Scheme 1, **3**) are most often synthesized by extension of the method for synthesis of nucleoside triphosphates in which activated nucleoside mono-, or thiomonophosphates **1** are reacted with pyrophosphate, or methylene-bis-phosphonates **2**. If excess of **2** is used the reaction leads to mono-nucleoside triphosph(on)ates. If a limiting amount of **2** is used, the reaction gives bis-nucleoside tetraphosph(on)ates **3**, although in low yield rarely exceeding 30%, together with significant amount of byproducts, mainly mono-, or bis-nucleoside polyphosphates with variable chain length. For instance, reaction of 2 equivalents of adenosine monophosphate (AMP), activated as the phosphoromorpholidate (**1**, X = 1-morpholinyl, Y = O, B = adenine) with one equivalent of pyrophosphate (**2a**), gave Ap₄A, 23%, together with bis-adenosine



Scheme 1 Synthesis of bis-nucleoside tetraphosph(on)ates from activated nucleoside monophosph(on)ates.

diphosphate (Ap₂A), 8%, triphosphate (Ap₃A), 18%, and pentaphosphate (Ap₅A), 4%, along with adenosine 5'-triphosphate (ATP), 7%; adenosine 5'-tetraphosphate, 8%; and minor amounts of AMP and adenosine 5'-diphosphate (ADP).¹⁸ Activated nucleoside monophosphates **1** have included morpholidates,¹⁹ imidazolidates,²⁰ and diphenylphosphoryl anhydrides.²¹

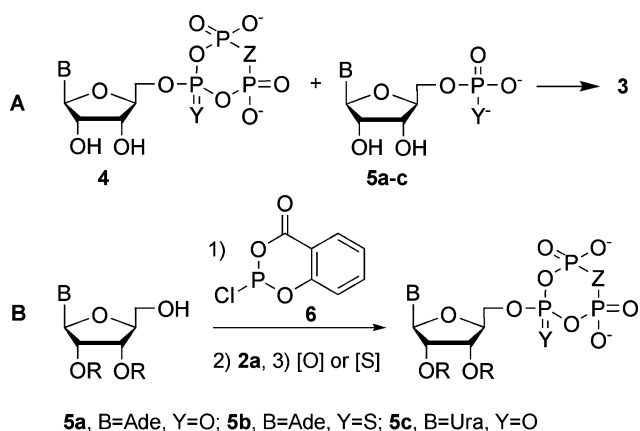
The need for activation of the nucleoside mono- or thiomonophosphates is a definite disadvantage of this approach, as it is often a source of side reactions.²²

In a modification of this method the activated nucleoside monophosphate **1** is reacted with nucleoside triphosphates or analogs to prepare bis-nucleoside tetraphosphates. For instance, the P^2, P^3 -dichloromethylene analog of Ap₄A (**3**, Y = O, Z = CCl₂) was prepared in 46% yield from the morpholidate of AMP (**1**, X = 1-morpholinyl, Y = O, B = adenine) and the P^2, P^3 -dichloromethylene analog of ATP, AppCCl₂p.¹⁹ In this method the higher yield is offset by the need to prepare the corresponding nucleoside triphosphates.

Another approach to the synthesis of bis-nucleoside tetraphosphates is based on the reaction of a P^1, P^3 -cyclic nucleoside triphosph(on)ates (“nucleoside trimetaphosphates”, Scheme 2A,

GLSynthesis Inc., One Innovation Dr, Worcester, MA, 01605, USA. E-mail: ivan.yanachkov@glsynthesis.com; Fax: +1 508 754 7075; Tel: +1 508 754 6700

† Electronic supplementary information (ESI) available: Spectroscopic and chromatographic data for the synthesized compounds. See DOI: 10.1039/c0ob00542h

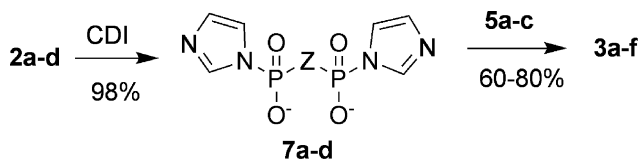


Scheme 2 Synthesis of nucleoside bis-tetraphosph(on)ates from nucleoside P^1, P^3 -cyclic triphosph(on)ates.

4) with a nucleoside monophosphates or thiomonophosphates (5). The nucleoside trimetaphosphates 4 can be prepared by treatment of the corresponding nucleoside triphosphates with carbodiimides^{14,23} or by phosphitylation of the 5'-hydroxy group of 2',3'-protected nucleosides with the salicyl phosphoanhydride reagent 6 (Scheme 2B), followed by double displacement of the salicylate with pyrophosph(on)ates (2) and oxidation or sulfurization of the resulting cyclic phosphite to 2',3'-protected 4.²⁴ These methods require the availability or the synthesis of the corresponding nucleoside triphosph(on)ates or protected nucleosides.

In yet another approach Tarussova *et al.*²⁵ reacted ADP with carbonylditriazole or carbonyldibenzimidazole to prepare Ap_4A in 16%, or 21% yield, respectively, presumably by formation of P^2 -imidazolide of ADP, and its condensation with unreacted ADP. The same activating agents, as well as carbonyldi-(4(5)-bromoimidazole) were used in the same work, and 1,1'-carbonyldiimidazole (CDI) was used in a previous work²⁶ to activate methylene-bis-phosphonic acid, which was then reacted with AMP to afford the P^2, P^3 -methylene analogue of Ap_4A in 5 to 40% yield.

In our search for a more efficient and cost-effective way to prepare bis-nucleoside tetraphosph(on)ates 3, we found that organic salts of pyrophosphoric acid and its halomethylene-bis-phosphonate analogs 2 react with excess CDI to give stable, isolable diimidazolides (Scheme 3, 7), and that these diimida-



- 7a**, Z=O; **7b**, Z=CHCl; **7c**, Z=CHF, **7d**, Z=CCl₂
3a, B=Ade, Y=Z=O, Ap_4A
3b, B=Ade, Y=S, Z=O, $Ap(S)ppp(S)A$
3c, B=Ade, Y=O, Z=CHCl, $AppCHClppA$
3d, B=Ade, Y=S, Z=CHCl, $Ap(S)ppCHClpp(S)A$
3e, B=Ade, Y=O, Z=CHF, $AppCHFppA$
3f, B=Ura, Y=Z=O, Up_4U

Scheme 3 Synthesis of bis-imidazolides of pyrophosphoric and halomethylene-bis-phosphonic acids 7 and their use for synthesis of bis-nucleoside tetraphosph(on)ates 3.

zolides react with nucleoside 5'-mono- or thiomonophosphates 5 to give bis-nucleoside tetraphosph(on)ates 3 directly and in high yields (Scheme 3). We present studies on the mechanism of formation of the novel diimidazolides 7, their properties and stability, and their utility in efficient, high yield synthesis of bis-nucleoside tetraphosph(on)ates.

Results and discussion

Synthesis of the diimidazolides of pyrophosphate and halomethylene-bis-phosphonates

If excess CDI is added to a solution of tetrabutylammonium pyrophosphate in DMF release of carbon dioxide is observed, and, at the end of this process, ³¹P NMR of the reaction mixture reveals that the pyrophosphate signal is replaced by a singlet at -21.26 ppm (Fig. 1). This reaction product was isolated by addition of 2 M NaClO₄ in acetone to the reaction mixture followed by further dilution with acetone. This resulted in a fine precipitate which was collected by centrifugation and washed repeatedly with acetone. This product was shown by ¹H, ³¹P, ¹³C NMR and ESI-MS to be the sodium salt of the P^1, P^2 -diimidazolide of pyrophosphoric acid (7a). It was obtained in nearly quantitative yield and in high purity (>97% by ³¹P NMR).

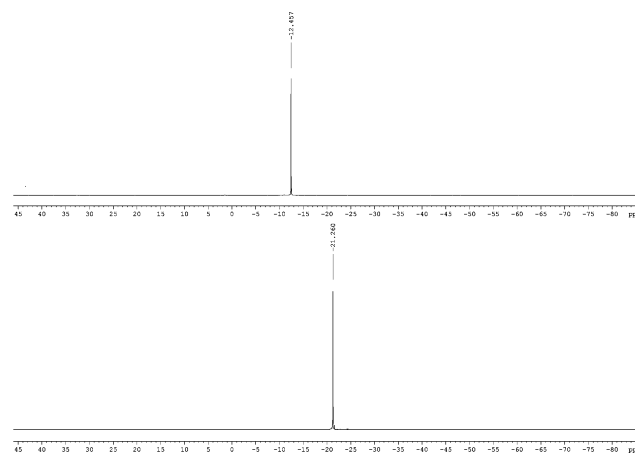


Fig. 1 ³¹P NMR of the reaction of tetrabutylammonium pyrophosphate with CDI in DMF. Top, before addition of CDI; bottom, after completion of the reaction.

Using the above procedure, we also prepared the diimidazolides of monochloromethylene-bis-phosphonate (7b), monofluoromethylene-bis-phosphonate (7c), and dichloromethylene-bis-phosphonate (7d). As with 7a, these compounds were isolated in almost quantitative yields and high purity as the disodium salts and characterized by their ¹H, ³¹P, ¹³C, and MS spectra.

It should be noted that the rate of the reaction is strongly dependent on the acidity/basicity of the reaction medium. For instance, it took 24 h for compound 2b as the mono-pyridinium salt to be converted into compound 7b when reacted with 5 mol mol⁻¹ CDI in DMF, and the final product was only of 94% purity (³¹P NMR), whereas, the same conversion took only 45 min when additionally 3 mol of triethylamine were present, and the purity of 7b was above 98%. Similarly, mono-tetrabutylammonium pyrophosphate reacted with a large excess of CDI very slowly, giving significant amounts of by-products resulting from cleavage of the

Table 1 Phosphorus chemical shifts and coupling constants of the intermediates in Scheme 4

Cpd. No.	Chemical shifts ^a (ppm)		Coupling constants (Hz)	
	P _a	P _b	J _{P_a-P_b}	J _{P-C}
8	-9.93	-19.91	22.4	4.2
9	-18.99	—	—	5.9
10	-21.62	—	—	2.1
11	-9.56	-19.64	19.4	—
12	-21.28	-20.95	23.8	4.2

^a For compound **12** from simulated fitting of an AB system. For all other compounds directly measured from spectra.

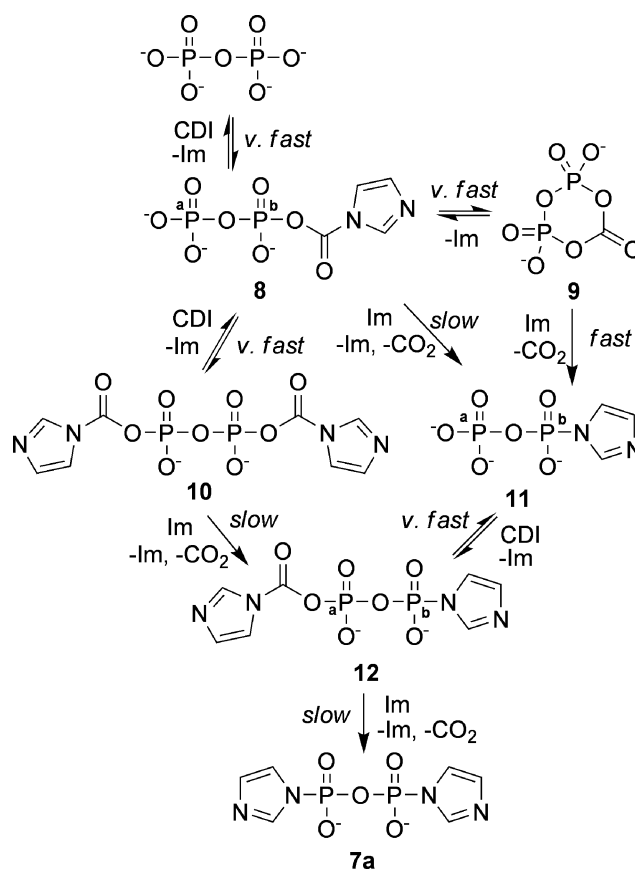
pyrophosphate bond, whereas, after addition of 3 equivalents of triethylamine, the reaction with 3 equivalent of CDI was very fast, giving almost exclusively **7a** (³¹P NMR).

Mechanism of formation of the diimidazolides

The high selectivity of the formation of the diimidazolides and the lack of any significant amount of by-products was quite surprising, having in mind that pyrophosphate and its analogs are bi-functional compounds. The formation of the diimidazolides is not instantaneous, and intermediate mono-imidazolides could self-condense, resulting in formation of polyphosphate by-products. To shed light on this unexpected high selectivity we studied the kinetics of this reaction by ³¹P NMR²⁷ in DMF-d₇. When CDI and tetrakis-tributylammonium pyrophosphate were reacted in the molar ratio 1.1 : 1, the ³¹P NMR of the reaction mixture revealed a complex and dynamic mixture of pyrophosphate derivatives (Fig. 2, A). When the reaction was carried out with an excess of CDI (molar ratio 3 : 1), significantly simpler spectra were obtained, in which two intermediates gradually converted into the final reaction product, the bis-imidazolide **7a**, represented by the singlet at -20.8 ppm (Fig. 2, B)

To assign the structures of the intermediates we carried out the above reaction using CDI labeled with ¹³C on the carbonyl group (¹³C-CDI). We used the 2-bond ¹³C-³¹P coupling to assign the ³¹P resonances of intermediates that contained the -P-O-¹³C(O)- mixed anhydride groups. Additionally, the resonances arising from phosphorus directly connected to imidazole (P-Im groups) were assigned thanks to the rather small 3-bond ¹H-³¹P coupling through the nitrogen, which resulted in broadening of those signals in the proton-coupled ³¹P spectra, when compared with the corresponding proton-decoupled spectra. Using this technique, as well as the strong 2-bond POP coupling in the non-symmetric structures and the information from the chemical shifts and the relative intensities, we were able to assign the structures of all intermediates observed in the ³¹P spectra. Those structures are shown in Scheme 4, and the assignments of their corresponding ³¹P resonances are given in Fig. 2A, spectrum 3 and listed in Table 1. The relative concentrations of each intermediate at different time points were determined by integration of the ³¹P spectra, and are plotted in Fig. 3 (panel A for pyrophosphate/CDI ratio 1 : 1.1 and panel B for ratio 1 : 3).

Based on the structural assignments and the rate of formation and consumption of the intermediates, a mechanism of the reaction of pyrophosphate with CDI, leading to **7a** was proposed, and is presented in Scheme 4.



Scheme 4 Mechanism of the reaction of CDI with pyrophosphate.

First, CDI reacts quickly with the primary phosphate groups of PP_i to form the mixed anhydrides **8** and **10**. Because of the high rate of this reaction, only intermediate **10** (together with a small amount of the products of its reaction with imidazole, **12** and **7a**) and no intermediate **8** is observed after 5 min in the reaction of PP_i with excess CDI (Fig. 2B, spectrum 3). This very fast conversion of the primary phosphate groups to mixed anhydrides explains why no by-products arising from self condensation of PP_i are observed when excess of CDI is employed, while they are present and gradually increase with time when equimolar amount of CDI is used (indicated with “*” on Fig. 2A). Second, the mixed anhydrides **8**, **10**, and **12** react with imidazole to form the imidazolides **11**, **12**, and **7a**, respectively. Here we observed significant difference in the rate of reaction of the mixed anhydride **8**, which has a neighboring primary phosphate group, and the mixed anhydrides **10** and **12**, in which the neighboring phosphate group is di-substituted. Analysis of the logarithmic plots of the decay portions of the reactions (Fig. 3, plot C) revealed that compound **8** is consumed much faster than compounds **10** and **12**. The reactions of imidazolysis of the mixed anhydrides **8**, **10**, and **12** appear to be of first overall order, which is not surprising if we take into account that imidazole is both consumed and released in these reactions, resulting in zero net imidazole concentration change. In the preceding reactions of formation of the mixed anhydrides the net imidazole concentration increases, but they are much faster to influence the order of the following reactions of imidazolysis. The first order rate constant of imidazolysis of compound **8** in the experiment with 1 : 1.1 molar ratios is 0.199 min⁻¹ (*t*_{1/2} = 3.47 min),

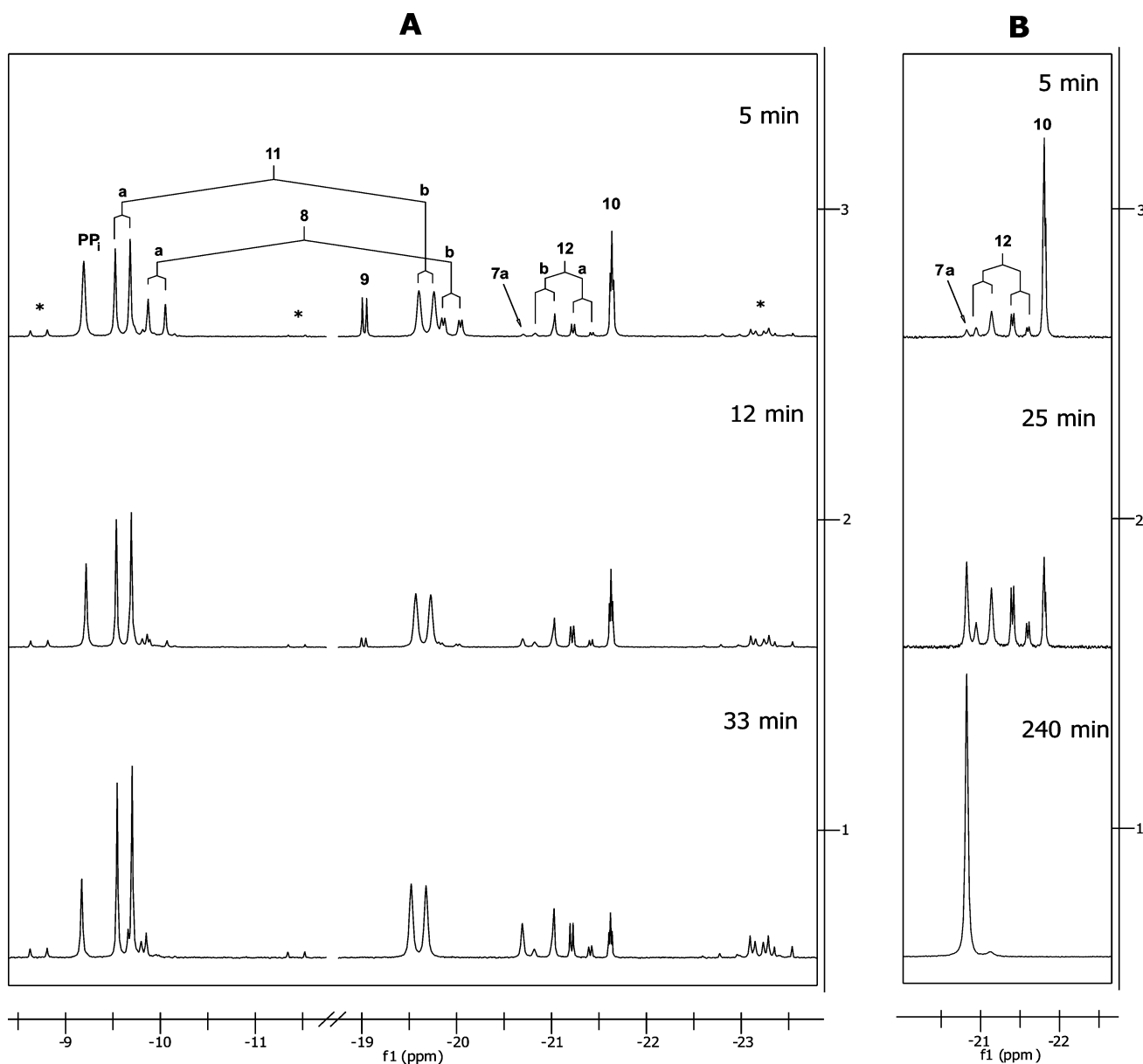


Fig. 2 ^{31}P NMR spectra (proton-coupled) of the time course of reaction of tetrakis-tributylammonium pyrophosphate, 0.16 M, in DMF-d_7 with ^{13}C -labeled CDI. Panel A: 1.1 mol CDI per mol pyrophosphate; from top to bottom: 5, 12 and 33 min after CDI addition. Panel B: 3 mol CDI per mol pyrophosphate, from top to bottom: 5, 25 and 240 min after CDI addition. “*” Designates the condensation by-products (tetraphosphates).

whereas the corresponding rate constant for compound **10** is 0.042 min^{-1} ($t_{1/2} = 16.5 \text{ min}$). The decay portion of the concentration–time curve of compound **12** was not observed in this experiment, but was clearly observed in the experiment with excess of CDI, where it corresponds to a pseudo-first-order process with a rate constant of 0.023 min^{-1} ($t_{1/2} = 30.1 \text{ min}$). In the same experiment the corresponding rate constant and half-life of compound **10** were 0.062 min^{-1} , and 11.2 min, respectively. The higher reaction rate for this compound in the experiment with excess of CDI is a result of the higher overall imidazole concentration.

While the higher reactivity of **8** in comparison with **10** and **12** can be attributed to an intramolecular general base catalysis by the doubly charged primary phosphate group, interacting either with the attacking imidazole, or with the leaving group, the observation

of the doublet centered at -18.99 ppm in the ^{31}P spectrum (Fig. 2A, spectrum 3), assigned to the cyclic mixed anhydride **9** ($^3J_{\text{C-P}} = 5.9 \text{ Hz}$) in Scheme 4, and the fact that the rate of decay of **9** equals that of compound **8** ($k = 0.195 \text{ min}^{-1}$ for compound **9**, and 0.199 min^{-1} for compound **8**) make the mechanism proposed in Scheme 3 more likely. According to this mechanism, compound **8** undergoes very fast intramolecular nucleophilic displacement at the carbonyl group to give **9**. A nucleophilic attack by imidazole on the carbonyl group of compound **9** results in formation of compound **8**, thus establishing very fast equilibrium between the two compounds with a equilibrium constant of 0.25, estimated from the ^{31}P NMR data. Thus, compound **11** can be formed by a nucleophilic displacement by imidazole at the phosphorus atom of compound **9**, followed by decarboxylation,

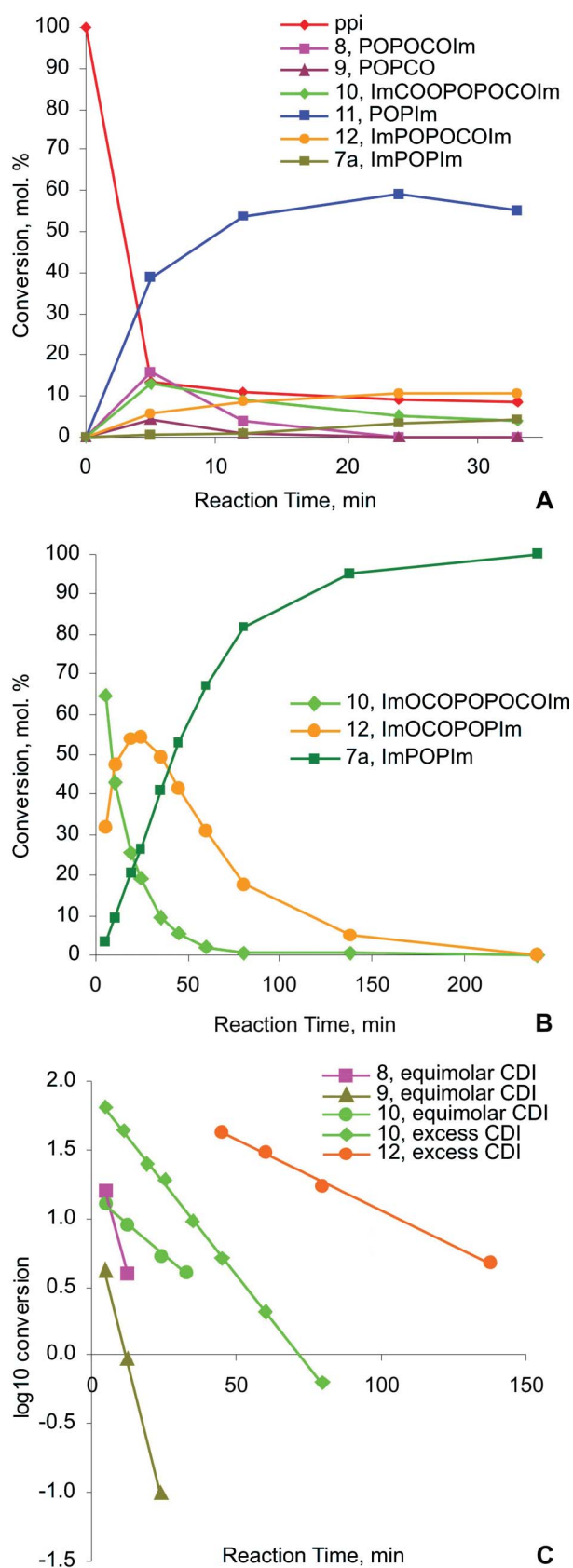


Fig. 3 Time course of the reaction of tetrakis-tributylammonium pyrophosphate, 0.16 M, with CDI. A: 1.1 mol CDI per mol pyrophosphate; B: 3 mol CDI per mol pyrophosphate; C: $\log_{10}(\text{conversion})$ –time plot.

or directly from compound **8**. Based on the ratio of the pseudo-first-order rate constants of decay of compounds **10**, **8** and **9**, and the equilibrium constant for interconversion of **8** and **9**, we estimated that compound **9** reacts with imidazole 20 times faster than compound **8**. To our knowledge this is the first observation of the cyclic anhydride **9**, and further studies of its reactivity/stability are under way.

Properties and stability of diimidazolides of diphosph(on)ates

The disodium salts of diimidazolides of pyrophosphoric and halomethylene-bis-phosphonic acids (**7a–d**) are white, non-hygroscopic powders, which when protected from atmospheric moisture are stable at room temperature for at least 8 months. They gradually (within a couple of days) decompose when exposed to moist air. They have excellent solubility in water, moderate solubility in methanol, low solubility in DMF, and are not soluble in acetone and other less polar organic solvents. As the bis-triethylammonium, -tributylammonium, or -tetrabutylammonium salts they are viscous oils, which are very soluble in anhydrous DMF and DMSO. The resulting solutions are quite stable – storage under Ar at room temperature for one week, and for months in the refrigerator, did not result in any signs of degradation (^{31}P NMR).

In an attempt to prepare the internally protonated, zwitterion form of **7a**, we subjected its triethylammonium salt to repeated evaporations from DMF and then from DMSO at high vacuum and room temperature. This resulted only in products of its self-condensation and degradation (^{31}P NMR). On the other hand, the non-volatile bis-tetrabutylammonium salt was completely stable under the same treatment, which is an indication that the internally protonated, zwitterion form is highly reactive and unstable.

The diimidazolides are remarkably stable in water at high pH. For instance, compound **7a** remained unchanged in 50 mM sodium carbonate/bicarbonate buffer at pH 9.8 for 48 h at room temperature (^{31}P NMR). At pH 7.8 in 50 mM sodium bicarbonate buffer *ca.* 50% of it decomposed after 19 h (determined by integration of ^{31}P NMR spectrum), resulting in a mixture of mono-imidazolidine **11**, 16.4%; PP_i , 34.9%; and **7a**, 48.7%. This result indicates that the diimidazolides are more stable hydrolytically than the mono-imidazolides, which could be attributed either to an intramolecular catalysis by the primary phosphate group of the mono-imidazolides; or by a catalytic effect of the bicarbonate anion, involving as reactive intermediate compound **9**. At pH lower than 7 the decomposition was too fast to be kinetically observed by phosphorus NMR.

Use of the diimidazolides of diphosph(on)ates for synthesis of bis-nucleoside tetraphosph(on)ates

Compounds **7a–c**, as the tri- or tetrabutylammonium salts, react with nucleoside mono- or thiomonophosphates **5a–c** at room temperature in concentrated DMF solutions to give the corresponding bis-nucleoside tetraphosph(on)ates **3a–f**. For instance the diimidazolidine **7b** reacted with the trioctylammonium salt of AMP for 30 h at room temperature in DMF to give 70% AppCHClppA (**3c**, isolated yield). It is important to keep the reaction concentration high: below 0.1 M the reaction rates become unacceptably low.

This reaction is catalyzed by 1*H*-tetrazole, pyridinium hydrochloride, and, most effectively, anhydrous zinc chloride.²⁸ The latter catalyst not only shortened the reaction times from 24–48 h to 0.5–2 h, but also, when used in sufficiently large excess (5–10 mol per mol diimidazolide) rendered the sodium salts of the diimidazolides **7** soluble in anhydrous DMF. This was a significant improvement, because the sodium salts, being non-hygroscopic powders, are much easier to handle than the tributylammonium salts, which are hygroscopic oils or glassy residues. The zinc chloride catalyst also made possible the use of the sodium salts of the nucleoside mono- or thiomonophosphates instead of their tributylammonium salts.²⁸ After completion of the reaction the zinc was sequestered by treatment with CHELEX[®] ion exchange resin in the sodium form or with EDTA, and the product was isolated by either ion-exchange chromatography on QEA-Sephadex[®] with a gradient of triethylammonium bicarbonate (TEAB) or reverse-phase chromatography with a gradient of acetonitrile in TEAB. After removal of the volatile buffer the products were converted into the sodium salts by either precipitation with sodium perchlorate/acetone, or by passing through strong cationite (in the sodium form) and lyophilization. Thus a series of bis-nucleoside tetraphosph(on)ates **3a–f** was prepared in yields in the range of 60–70% for the pyrophosphate analogs and 70–80% for the halomethylene-bis-phosphonate analogs, and were characterized by ¹H, ³¹P NMR and UV spectra and LC–MS in the positive and negative ionization modes.

Experimental

General

Chloromethylene-bis-phosphonic acid, **2b**,²⁹ dichloromethylene-bis-phosphonic acid, **2d**,³⁰ fluoromethylene-bis-phosphonic acid, **2c**,³¹ and 1,1'-carbonyldiimidazole labeled with ¹³C on the carbonyl group,³² were prepared by the corresponding literature methods. All transfers and manipulations at moisture-sensitive steps were done under argon, using syringe techniques and oven-dried and argon-filled glassware. The purity of the synthesized bis-nucleoside tetraphosph(on)ates was determined by analytical reverse-phase HPLC (column, XBridge Shield C18; mobile phases – A, 20 mM triethylammonium acetate in water pH 6.8; B, acetonitrile; linear gradient from 0 to 30% B in A for 30 min; UV detection at 260 nm) and was above 95%.

Solutions of tributylammonium pyrophosphate and tetrabutylammonium pyrophosphate in DMF

Tetrasodium pyrophosphate decahydrate (10.00 g, 22.4 mmol) was dissolved in 100 ml water. This solution was passed through a column (20 × 6.3 cm) of Dowex[®] 50X4-100 in the H⁺ form, and eluted with ice-cold water until the eluate pH increased to 5. The eluate was collected into a cooled (ice bath) and stirred solution of tri-*n*-butylamine (8.31 g, 10.7 ml, 44.8 mmol) in 200 ml of 2-propanol. The resulting solution was concentrated under vacuum (bath temperature 35 °C), and then re-evaporated under high vacuum (less than 1 torr) twice from 150 ml DMF. The resulting oil was transferred quantitatively with 20 ml of dry (SureSeal[®]) DMF into a pre-tared flask, evaporated under high vacuum, dissolved in 20 ml dry DMF, and partially evaporated to give 27.93 g tributylammonium pyrophosphate solution in DMF with concentration of 0.80 mmol g⁻¹ solution.

To prepare tetrabutylammonium pyrophosphate a 55% w/w tetrabutylammonium hydroxide solution in water (21.8 ml, 21.15 g, 44.8 mmol) was added with stirring and cooling to the ice-cooled eluate of pyrophosphoric acid from the ion-exchange column, and the resulting solution was evaporated and rendered anhydrous as described for the tributylammonium salt to obtain 46.5 g of DMF solution at a concentration 0.482 mmol g⁻¹. Both the tributylammonium and the tetrabutylammonium salt solutions were sealed under argon and stored at –5 to –10 °C. After 1 year, ³¹P NMR of the tributylammonium salt solution showed less than 1 mol% disproportionation to ortho- and tripolyphosphate.

Monitoring the reaction of tributylammonium pyrophosphate with 1,1'-¹³C-carbonyldiimidazole by ³¹P-NMR

Tributylammonium pyrophosphate (0.175 g of 0.80 mmol g⁻¹ solution, 0.144 mmol, measured by means of a pre-tared syringe) was mixed with 0.6 ml DMF-d₇, and evaporated on a speed-vac to 0.2 ml. After breaking the vacuum with argon the residue was mixed with a solution of ¹³C-CDI (25 mg, 0.154 mmol) and tributylamine (71 μl, 52 mg, 0.28 mmol) in 0.6 ml DMF-d₇. The mixture was transferred into a 5 mm NMR tube, and ³¹P spectra (32 scans each) were recorded at 5, 12, 24, and 33 min at 25 °C. To improve the integration accuracy, long (5 s) acquisition times without proton decoupling were used.

In a separate experiment 75 mg (0.46 mmol) of ¹³C-CDI was used, and the spectra were recorded at 5, 11, 19, 25, 35, 45, 60, 80, 138, and 240 min.

P¹,P²-Di(1-imidazolyl)pyrophosphate, **7a**

N,N'-Carbonyldiimidazole (2.43 g, 15.0 mmol) was suspended in dry DMF (5 ml) under argon. The flask was sealed with a septum, connected with a mineral oil bubbler, and tributylammonium pyrophosphate (6.25 g of 0.80 mmol g⁻¹ solution in DMF, 5.0 mmol) was added by means of pre-tared syringe with stirring, followed by tributylamine (0.927 g, 1.19 ml, 5.0 mmol). The mixture was stirred overnight at r.t., during which time carbon dioxide evolution ceased. Water (200 μl) was added and the mixture was stirred for 5 min, and then concentrated under 0.5 mm Hg vacuum at 32 °C to an oil. This oil was taken up in 50 ml dry DMF and concentrated partially as above to give 15.4 g of DMF solution of **7a** as the tributylammonium salt. This solution contains 0.324 mmol g⁻¹ **7a** and 1.30 mmol g⁻¹ imidazole, and can be used for synthesis of bis-nucleoside tetraphosph(on)ates, as in the example for synthesis of **3a** below without farther purification or imidazole removal.

Pure disodium salt of **7a** was isolated as follows: The DMF solution from above was evaporated under vacuum (0.5 mm Hg, 32 °C) to oil. This oil was added with vigorous stirring to 250 ml of 2 M solution of sodium perchlorate in acetone, followed by 250 ml acetone. The mixture was stirred vigorously for 30 min, during which time the initially formed sticky residue converted into a white fine powder. This powder was separated by centrifugation and washed twice by resuspension in 100 ml acetone, centrifugation and decanting. After initial drying under a stream of nitrogen the product was dried under high vacuum for 16 h. Yield, 1.58 g, 98%. ¹H NMR (300 MHz, D₂O) δ 7.64 (s, 1H), 7.06 (s, 1H), 6.94 (s, 1H); ³¹P NMR (D₂O) –20.87 (s); ¹³C

NMR (D₂O) 140.28, 129.36 (t, ³J_{PC} = 5.1 Hz), 120.6; MS (ESI in the negative ionization mode): calcd. for C₆H₇N₄O₅P₂⁻ (M - H) 276.99 (100%), 277.99 (8.2%), 278.99 (1.1%); found 276.97 (100%), 277.97 (7.4%), 278.96 (1.0%); positive ionization mode: calcd. for C₆H₈N₄NaO₅P₂⁺ (M + Na) 300.99 (100%), 301.99 (6.8%), 301.98 (1.5%); found 300.86 (100%), 301.89 (7.9%), 302.88 (1.3%).

Disodium chloromethylene-bis(1-imidazolyl)phosphonate, 7b

Monochloromethylene-bis-phosphonic acid mono-pyridinium salt (500 mg, 1.173 mmol), CDI (840 mg, 5.18 mmol), and dry DMF were mixed under argon in a dry vial equipped with a stirring bar. The vial was sealed, equipped with a bubbler, and triethylamine (725 μl, 524 mg, 5.18 mmol) was added *via* a syringe with stirring. After a few min the mixture became homogenous. After 4 h sodium perchlorate (5 ml of 2 M solution in acetone) was added with vigorous stirring, followed by acetone (15 ml). The mixture was stirred vigorously for 2 h, and then centrifuged. The supernatant was discarded, and the solid was suspended in anhydrous acetone (30 ml). The suspension was centrifuged, the supernatant discarded, and the acetone washing was repeated two more times. After the final wash the solid was dried under a stream of dry nitrogen and then under high vacuum at r.t. for 24 h to give 610 mg (99.6%) of white powder. ¹H NMR (300 MHz, DMSO-d₆) δ 7.66 (2H, bs, H-2), 7.15 (2H, bs, H-4), 6.80 (2H, bs, H-5), 3.69 (1H, t, CH-Cl, ²J_{P-H} = 15.48 Hz); ³¹P NMR (121 MHz, DMSO-d₆), ppm: 0.63 (¹H decoupled, s; ¹H coupled, d, ²J_{P-H} = 14.92 Hz); MS (ESI in the negative ionization mode): Calcd. for C₇H₈ClN₄O₄P₂⁻ (M - H), 309.0 (100%), 310.0 (9.0%), 311.0 (32%); Observed, 309.1 (100%), 310.0 (6.5%), 311.1 (35%)

Tetrasodium P¹,P⁴-bis(adenosine-5′)-P²,P³-(chloromethylene)-tetraphosphate, 3c (uncatalyzed reaction)

Adenosine 5′-monophosphate, free acid, monohydrate (1.09 g, 2.98 mmol), tri-octylamine (1.055 g, 2.98 mmol, 1.31 ml), and methanol (50 ml) were stirred until a clear solution was obtained. The solution was evaporated (30 °C, 10–15 mm Hg vacuum). The residue was dissolved in 100 ml dry DMF, and the solution was evaporated (30 °C, 0.5–1 mm Hg vacuum). The residue was re-dissolved in another 100 ml of dry DMF, and the evaporation was repeated under the same conditions.

Chloromethylene-bis-phosphonic acid, mono-pyridinium salt (216 mg, 0.746 mmol) and tributylamine (277 mg, 1.49 mmol, 0.355 ml) were stirred with 15 ml methanol until a clear solution was obtained (*ca.* 15 min.). This solution was evaporated (30 °C, 10–15 mm Hg vacuum) to a glassy residue. This residue was dissolved in 50 ml dry DMF, and evaporated (30 °C, 0.5–1 mm Hg vacuum) to an oily residue. The residue was re-dissolved in another 50 ml of dry DMF, and the evaporation was repeated under the same conditions. The resulting chloromethylene-bis-phosphonate, bis-tributylammonium salt, was dissolved in 5 ml dry DMF under argon, and CDI (0.605 g, 3.73 mmol) was added in one portion with stirring. The flask was sealed and connected to a bubbler filled with paraffin oil. The rate and the advance of the reaction was monitored by the rate of release of CO₂. (After 12 h a small portion of the reaction mixture was diluted with DMF-d₇ and checked by ³¹P NMR. No starting material was present, and only one phosphorus signal, corresponding to the diimidazolide was observed).

To decompose excess CDI, water (100 μl) was added. After 5 min the reaction mixture was concentrated to 2/3 of its volume (30 °C, 0.5–1 mm) and mixed with the solution of AMP, trioctylammonium salt, prepared above. The mixture was concentrated again (30 °C, 0.5–1 mm Hg vacuum) to a light oil. The vacuum was broken with argon, and the reaction mixture was stirred under argon for 36 h at r.t.. Water (50 ml) was added, and the mixture was extracted twice with 150 ml diethyl ether containing 1% triethylamine. The aqueous layer was evaporated under vacuum to half of its volume, and then was loaded on a Toyopearl DEAE-650M column (5 × 45 cm) which was pre-equilibrated with 0.2 M triethylammonium bicarbonate (TEAB) buffer, pH 8, containing 10% v/v acetonitrile. The elution was carried out with a gradient from the equilibration buffer to 1 M TEAB/acetonitrile, 9:1 v/v, for 300 min at a flow rate of 20 ml min⁻¹. The fractions containing the product were pooled and evaporated (35 °C, 10–15 mm Hg vacuum) with periodic addition of 1-butanol to prevent foaming. The residue was evaporated 3 times from methanol (100 ml each) under vacuum and then dried for 4 h at 0.2 – 1 mm Hg vacuum in order to remove any residual TEAB, and then dissolved in methanol (5 ml). A solution of sodium perchlorate (1 g) in methanol (5 ml) was added dropwise with vigorous stirring. Acetone (50 ml) was added with stirring, and after 30 min the mixture was centrifuged, and the clear supernatant was decanted. The white solid was re-suspended in 10 ml acetone, and the suspension was filtered through a 0.45 μm glass fiber filter. The solid was washed with three portions of acetone (5 ml each), and with 5 ml diethyl ether, and dried under a stream of nitrogen, and then under high vacuum to give 502 mg (70%) of **3c** as a fine white powder. ¹H NMR (300 MHz, D₂O) δ 8.32, 8.30 (2H, s, H-8), 8.01 (2H, bs, H-2), 5.86 (2H, d, H-1′, ³J_{1′-2′} = 7.48 Hz), 4.46 (2H, m, H-2′), 4.37 (2H, m, H-3′), 4.30 (2H, m, H-4′), 4.25 (1H, t, CHCl, ²J_{P-H} = 17.1 Hz), 4.09–4.24 (4H, m, H-5′,5′′); ³¹P NMR (121 MHz, D₂O), ppm: 5.03 (m, P² + P³, ²J_{P¹-P²} = 25.6 Hz), -8.02 (m, P¹ + P⁴, ²J_{P¹-P²} = 25.5 Hz, ³J_{P¹-P³} = 8.5 Hz); MS (ESI in the negative ionization mode): Calcd. for C₂₁H₂₈ClN₁₀O₁₈P₄⁻ (M - H), 867.0 (100%), 868.0 (27.4%), 869.0 (39.3%), 870.0 (10.1%), 871.0 (2.6%); Observed, 867.1 (100%), 868.1 (27.6%), 869.0 (35.2%), 870.0 (9.4%), 871.0 (2.3%).}}}}

Tetrasodium P¹,P⁴-bis(adenosine-5′)-P²,P³-(fluoromethylene)tetraphosphate, 3e (uncatalyzed reaction)

Trimethylbromosilane, 0.887 g, 0.767 ml, 5.8 mmol, was added dropwise under nitrogen and with stirring, to an ice cooled solution of tetraethyl (fluoromethylene)-bis-phosphonate²⁸ (306 mg, 1 mmol) in 1.5 ml dichloromethane. After 24 h at r.t. the mixture was evaporated under vacuum (45 °C, 10–15 mm Hg), re-evaporated 3 times from 30 ml methanol, and once from 10 ml dry DMF (33 °C, 0.5–1 mm Hg vacuum). The resulting oil was dissolved in a mixture of 10 ml dry DMF and 0.524 ml (2.2 mmol) tributylamine and evaporated again as above to give the bis-tributylammonium salt of **2c**. Following the procedure for compound **3c**, this salt was converted into the diimidazolide **7c** by reaction with CDI (0.811 g, 5 mmol) in 6.7 ml dry DMF. After decomposition of the excess of CDI with 130 μl water, **7c** was condensed with 4 mmol AMP, trioctylammonium salt, to give, after work up 941 mg (68%) of tetrasodium P¹,P⁴-bis(5′-adenosine)-P²,P³-(monofluoromethylene)tetraphosphate, **3e**. ¹H

NMR (300 MHz, D₂O) δ 8.27 (2H, s, H-8), 8.02 (2H, s, H-2), 5.92 (2H, d, H-1'), 5.07 (1H, dt, CHF, $^2J_{F-H} = 45.6$ Hz $^2J_{P-H} = 14.1$ Hz), 4.63 (2H, m, H-2'), 4.44 (2H, m, H-3'), 4.25 (2H, m, H-4'), 4.11 (4H, m, H-5', 5''); ^{31}P NMR (121 MHz, D₂O), ppm: 0.59–1.83 (m, P² + P³, $^2J_{P-F} = 75.0$ Hz), -9.61 – -10.30 (m, P¹ + P⁴); MS (ESI in the negative ionization mode): Calcd. for C₂₁H₂₈FN₁₀O₁₈P₄⁻ (M – H), 851.1 (100%), 852.1 (27.4%), 853.1 (7.3%), 854.1 (1.3%); Observed, 851.1 (100%), 852.1 (21.5%), 853.0 (4.5%), 854.2 (0.2%).

Tetrasodium P¹,P⁴-bis(adenosine-5')-tetrphosphate, **3a** (tetrazole catalysis)

Adenosine 5'-monophosphate, mono-tetrabutylammonium salt (3.81 g of DMF solution with concentration of 0.131 mmol g⁻¹, ³³ 0.500 mmol) and DMF solution of the tributylammonium salt of **7a**, containing 0.324 mmol **7a** and 1.30 mmol imidazole per gram (386 mg solution, 0.125 mmol **7a**) were mixed and concentrated under vacuum (33 °C, 0.5–1 mm Hg) to a volume of ca. 2 ml. A solution of 1*H*-tetrazole in acetonitrile (0.45 M, 1 ml, 0.45 mmol) was added with stirring under argon, which resulted in formation of a bulky white precipitate. This mixture was concentrated under vacuum (r.t., 0.5–1 mm Hg) to a final volume of ca. 1 ml. During this evaporation the precipitate dissolved, resulting in a clear oil. This oil was stirred for 16 h at r.t. under argon, and then diluted with 100 ml water. The solution was loaded on a 5 × 45 cm Toyopearl DEAE-650M column. The column elution and the product isolation and conversion to the tetra-sodium salt were carried out as described above for compound **3c** to give 85.0 mg (74%) of **3a** as a fine white powder. ¹H NMR (300 MHz, D₂O) δ 8.32 (2H, s, H-8), 8.09 (2H, s, H-2), 5.97 (2H, d, H-1', $^3J_{1'-2'} = 5.97$ Hz), 4.70–4.75 (1H, m, H-2'), 4.47–4.51 (2H, m, H-3'), 4.26–4.32 (2H, m, H-4'), 4.17–4.22 (4H, m, H-5', 5''); ^{31}P NMR (121 MHz, D₂O), ppm: -11.28 (m, P¹ + P⁴, $^2J_{P1-P2} = 17.0$ Hz, $^3J_{P1-P3} = 0.3$ Hz, $^4J_{P1-P4} = 0.7$ Hz, $^2J_{P2-P3} = 14.7$ Hz³⁴), -23.11 (m, P² + P³); MS (ESI in the negative ionization mode): Calcd. for C₂₀H₂₇N₁₀O₁₉P₄⁻ (M – H), 835.0 (100.0%), 836.0 (26.4%), 837.0 (7.2%), 838.0 (1.1%); Observed, 835.1 (100%), 836.1 (24.0%), 837.1 (6.7%), 838.1 (0.9%).

Tetrasodium P¹,P⁴-bis(uridine-5')-tetrphosphate, **3f** (zinc chloride catalysis)

Uridine 5'-monophosphate, disodium salt, hydrate (26% w/w water content, 274 mg, 0.576 mmol) was dissolved in 3 ml of water. This solution was cooled in ice and passed through a pre-cooled column (12 × 2 cm) of Dowex[®] 50X4-100 in the H⁺ form, and eluted with ice-cold water until pH of the eluate was above 5. The eluate was collected into a stirred mixture of triethylamine (0.25 ml), isopropanol (10 ml) and water (10 ml). The resulting solution was evaporated under vacuum (35 °C, 10–15 mm Hg), and the glassy residue was dissolved in anhydrous DMF (20 ml) and evaporated under vacuum (35 °C, 0.5–1 mm Hg). This triethylammonium salt was dissolved in 1.5 ml of dry DMF under argon. Disodium P¹,P²-di(1-imidazolyl)pyrophosphate, **7a** (67.1 mg, 0.208 mmol) followed by anhydrous zinc chloride (138 mg, 1.015 mmol) were added with rigorous stirring under argon. After 15 min a clear solution was obtained. After 3 h a cooled suspension of Chelex[®] resin in the sodium form (5.5 ml resin in 11 ml water) was added. The mixture was shaken for 15–20 min until a clear supernatant was obtained. The resin was

filtered and washed with water until the washings, when spotted on a TLC plate with fluorescent indicator, did not show a dark spot under UV light. The combined filtrate and washings were loaded on a column of DEAE Sephadex[®] (25 × 2.5 cm), which was equilibrated with 20 mM TEAB buffer, pH 8, containing 10% v/v acetonitrile. The elution was carried out with a linear gradient from the equilibration buffer to 1.5 M TEAB, pH 8, containing 10% acetonitrile for 800 min at 3 ml min⁻¹ flow rate. The fractions containing the product were pooled, evaporated, and converted into the sodium salt as described for compound **3c** to give 112 mg (61%) of **3f** as a fine white powder. ¹H NMR (300 MHz, D₂O) δ 7.75 (2H, d, H-6, $^3J_{H6-H5} = 7.91$ Hz), 5.90 (2H, d, H-1', $^3J_{1'-2'} = 5.28$ Hz), 5.80 (2H, d, H-5, $^3J_{H5-H6} = 7.72$ Hz), 4.21–4.31 (4H, m, H-2'+H-3'), 4.09–4.17 (6H, m, H-4'+H5', 5''); ^{31}P NMR (121 MHz, D₂O), ppm: -10.90 (m, P¹ + P⁴, $^2J_{P1-P2} = 17.7$ Hz, $^3J_{P1-P3} = 0.2$ Hz, $^4J_{P1-P4} = 1.8$ Hz), -22.34 (m, P²+P³, $^2J_{P2-P3} = 15.8$ Hz³⁴); MS (ESI in the negative ionization mode): Calcd. for C₁₈H₂₅N₄O₂₃P₄⁻ (M – H), 789.0 (100%), 790.0 (22.1%), 791.0 (7.1%), 792.0 (1.2%); Observed, 789.1, (100%), 790.0 (20.4%), 791.1 (6.5%), 792.1 (0.9%).

Tetrasodium P¹,P⁴-bis(adenosine-5')-P¹,P⁴-dithiotetraphosphate, **3b** (zinc chloride catalysis)

Adenosine 5'-thiomonophosphate bis-triethylammonium salt³⁵ (80 mg, 0.141 mmol) was evaporated twice from 1 ml dry DMF under vacuum (30 °C, 0.5–1 mm Hg). Disodium P¹,P²-di(1-imidazolyl)pyrophosphate, **7a** (11.0 mg, 0.035 mmol) and 0.4 ml dry DMF were added under argon. After 1 min sonification the mixture was evaporated as above. The vacuum was broken with argon, and the flask was sealed with a septum under argon. Anhydrous zinc chloride (96 mg, 0.707 mmol) was dissolved in 1 ml dry DMF. The solution was evaporated under vacuum (30 °C, 0.5–1 mm Hg), and re-dissolved under argon in 0.3 ml dry DMF. This solution was added by a syringe to the sealed flask with stirring at r.t. A clear solution quickly formed. After 2.5 h the reaction mixture was worked up with Chelex[®] resin, followed by chromatography on DEAE Sephadex[®] as described for compound **3f**. After conversion to the sodium salt as described for compound **3c**, using 1 ml of 2 M sodium perchlorate in acetone and 4 ml acetone, 3 washings with 5 ml acetone each, and drying under high vacuum, 22.5 mg of **3b** (67.5%) were obtained as a fine white powder. Because of the stereogenic nature of the phosphorothioate modifications at P¹ and P⁴ this compound is a mixture of 3 diastereomers with configurations R_P,R_P,S_P,S_P and R_P,S_{P} ≡ S_P,R_P (because of the molecular symmetry).^{36,37} These diastereomers were easily separable by analytical reverse-phase HPLC (see general methods) used to determine the product purity. The corresponding H-8 and H-2 resonances in the ¹H NMR spectra are resolved, while the phosphorus resonances in the ³¹P NMR spectrum are only partially resolved. ¹H NMR (300 MHz, D₂O) δ 8.460, 8.447, 8.351 (2H, s, three diastereomers H-8 in ratios 0.275 : 0.175 : 0.555); 8.039, 8.033, 8.029 (2H, three diastereomers H-2); 5.946 (2H, d, H-1', $^3J_{1'-2'} = 5.39$ Hz); 4.735–4.627 (2H, m, H-2'); 4.534–4.475 (2H, m, H-3'); 4.305–4.205 (6H, m, H-4'+H5', 5''); ^{31}P NMR (121 MHz, D₂O), ppm: 41.487–40.852 (overlapping AA'–XX' and AB–XY multiplets, P¹ + P⁴); -26.016 – -26.451 (overlapping AA'–XX' and AB–XY multiplets, P² + P³); MS (ESI in the negative ionization mode): Calcd. for C₂₀H₂₇N₁₀O₁₇P₄S₂⁻ (M – H), 867.0 (100.0%), 868.0 (27.9%), 869.0 (16.3%), 870.0}

(3.7%), 871.0 (1.1%); Observed, 867.2 (100%), 868.1 (25.9%), 869.1 (13.6%), 870.0 (2.8%), 871.0 (0.7%).

Tetrasodium P^1, P^4 -bis(adenosine-5')- P^2, P^3 -(chloromethylene)- P^1, P^4 -dithiotetraphosphate, **3d** (zinc chloride catalysis)

Compound **3d** was prepared using the procedure for compound **3b** but with **7b** instead of **7a**. The product was isolated in 72% yield. This analog also exists as a diastereomeric mixture (as for **3b**). The stereogenic chloromethyl group causes four diastereomers to exist, however.²¹ In fact, four peaks were observed under analytical reverse-phase HPLC (see general methods), and 7 peaks were observed for H-8 in the ¹H NMR spectrum. ¹H NMR (300 MHz, D₂O) δ 8.46–8.285 (2H, multiple singlets, H-8), 8.04–7.93 (2H, multiple singlets, H-2), 5.995–5.915 (2H, overlapping d, H-1'), 4.89–4.42 (1H, multiple overlapping triplets, CHCl), 4.72–4.59 (2H, m, H-2'), 4.53–4.46 (2H, m, H-3'), 4.365–4.28 (2H, m, H-4'), 4.28–4.115 (4H, m, H-5', 5''); ³¹P NMR (121 MHz, D₂O), ppm: 44.61–43.71 (overlapping AA', XX' multiplets, P¹ + P⁴), 3.45–2.90 (overlapping AA', XX' and AB, XY multiplets, P² + P³); MS (ESI in the negative ionization mode): Calcd. for C₂₁H₂₈ClN₁₀O₁₆P₄S₂⁻ (M – H), 899.0 (100.0%), 900.0 (28.9%), 901.0 (48.3%), 902.0 (13.1%), 903.0 (6.3%), 904.0 (1.4%); Observed, 899.0 (100%), 900.0 (28.7%), 900.9 (48.9%), 901.9 (11.6%), 903.0 (4.8%), 904.1 (0.5%).

Conclusions

Carbonyldiimidazole reacts with pyrophosphate and halomethylene-bis-phosphonates in DMF to form the corresponding P^1, P^2 -bis-imidazolides. The mechanism of this reaction involves fast formation of mixed anhydrides with imidazolyl-1-carbonic acid, which react slowly with imidazole to give the imidazolides. It appears that the cyclic anhydride between pyrophosphoric and carbonic acid is involved as an intermediate as well. The resulting bis-imidazolides when protected from moisture are stable, and can be isolated in excellent yields as the disodium salts which can be stored at room temperature. They hydrolyze very slowly at high pH, and are highly reactive and quickly decompose at low pH. These compounds, both as the pure sodium salts and as the salts formed *in situ* with organic amines or tertiary ammonium salts, are excellent reagents for the synthesis of bis-nucleoside tetraphosphates, and their P^1, P^4 -dithio-, and P^2, P^3 -halomethylene phosphonate analogs.

Acknowledgements

The authors are grateful to Dr Wei-Chu Xu for the synthesis of tetraethyl (fluoromethylene)-bis-phosphonate, a precursor for the synthesis of **2c**. This work was supported in part by SBIR grants HL081992 and HL088828 (to IBY) from the National Heart, Lung and Blood Institute.

References and notes

- 1 *Nucleoside triphosphates and their analogs: chemistry, biotechnology, and biological applications*, ed. M. Vaghefi, Taylor & Francis: Boca Raton, 2005.
- 2 K. Burgess and D Cook, *Chem. Rev.*, 2000, **100**(6), 2047–2060.
- 3 *Ap_nA and Other Dinucleoside Polyphosphates*, ed. A. G. McLennan, CRC Press: Boca Raton, 1992.
- 4 C. H. V. Hoyle, R. H. Hilderman, J. J. Pintor, H. Schlüter and B. F. King, *Drug Dev. Res.*, 2001, **52**, 260–273.

- 5 A. G. McLennan, *Pharmacol. Ther.*, 2000, **87**(2–3), 73–89.
- 6 H. Schlüter, E. Offers, G. Brüggemann, M. von der Giet, M. Tepel, E. Nordhoff, M. Karas, C. Spieker, H. Witzel and W. Zidek, *Nature*, 1994, **367**, 186–188.
- 7 Z. Palfi, G. Suranyi and G. Borbely, *Biochem. J.*, 1991, **276**, 487–491.
- 8 J. Eugen, G. Verspohl, M. H. N. Blackburn, J. Hagemann and M. Lempka, *J. Med. Chem.*, 2003, **46**, 1554–1562.
- 9 S. W. Chan, S. J. Gallo, B. K. Kim, M. J. Guo, G. M. Blackburn and P. C. Zamecnik, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 4034–4039.
- 10 J. Pintor, M. Diaz-Hernandez, J. Gualix, R. Gomez-Villafuertes, F. Hernando and M. T. Miras-Portugal, *Pharmacol. Ther.*, 2000, **87**(2–3), 103–115.
- 11 B. Walkowiak, J. Baraniak, C. S. Cierniewski and W. Stec, *Bioorg. Med. Chem. Lett.*, 2002, **12**(15), 1959–1962.
- 12 W. Pendergast, B. R. Yerxa, J. G. Douglass 3rd, S. R. Shaver, R. W. Dougherty, C. C. Redick, I. F. Sims and J. L. Rideout, *Bioorg. Med. Chem. Lett.*, 2001, **11**(2), 157–160.
- 13 E. A. Shirokova, A. L. Khandzhinskaya, Y. S. Skoblov, L. Y. Goryunova, R. S. Beabealashvili and A. A. Krayevsky, *Nucleosides, Nucleotides Nucleic Acids*, 2001, **20**(4), 1033–1036.
- 14 J. G. Douglass, R. I. Patel, B. R. Yerxa, S. R. Shaver, P. S. Watson, K. Bednarski, R. Flourde, C. C. Redick, K. K. Brubaker, A. C. Jones and J. L. Boyer, *J. Med. Chem.*, 2008, **51**(4), 1007–1025.
- 15 D. Kellerman, A. R. Mospan, J. Engels, A. Schaberg, J. Gorden and L. Smiley, *Pulm. Pharmacol. Ther.*, 2008, **21**(4), 600–607.
- 16 J. Tauber, W. F. Davitt, M. D. Bokoosky, K. K. Nichols, B. R. Yerxa, A. E. Schaberg, L. M. LaVange, M. C. Mills-Wilson and D. J. Kellerman, *Cornea*, 2004, **23**(8), 784–792.
- 17 Y. Kikuta, E. Ohiwa, K. Okada, A. Watanabe and S. Haruki, *Acta Anaesthesiol. Scand.*, 1999, **43**(1), 82–86.
- 18 D. L. M. Verheyden, W. E. Wehrl and J. G. Moffatt, *J. Org. Chem.*, 1965, **30**, 3381–3385.
- 19 G. M. Blackburn, M. J. Guo and A. G. McLennan, in *Ap_nA and Other Dinucleoside Polyphosphates*, ed. A. G. McLennan, CRC Press: Boca Raton, 1992, pp. 305–342.
- 20 N. B. Tarussova, T. I. Osipova, A. I. Biriukov, M. J. Pokrovskaya, C. V. Meshkov and N. V. Gnuchev, *Nucl. Acids Res. Symp. Ser.*, 1984, **14**, 287–288.
- 21 G. M. Blackburn and M. J. Guo, *Tetrahedron Lett.*, 1990, **31**, 4371–4374.
- 22 M. Maeda, A. D. Patel and A. Hampton, *Nucleic Acids Res.*, 1977, **4**(8), 2843–2853.
- 23 H. J. Ko, R. L. Carter, L. Cosyn, R. Petrelli, S. de Castro, P. Besada, Y. X. Zhou, L. Cappellacci, P. Franchetti, M. Grifantini, S. Van Calenbergh, T. K. Harden and K. A. Jacobson, *Bioorg. Med. Chem.*, 2008, **16**(12), 6319–6332.
- 24 Q. Han, B. L. Gaffney and R. A. Jones, *Org. Lett.*, 2006, **8**, 2075–2077.
- 25 N. B. Tarussova, T. I. Osipova, P. P. Purygin and I. A. Yakimova, *Bioorg. Khim.*, 1986, **12**, 404–407.
- 26 N. B. Tarussova, V. V. Shumiyanzeva, A. C. Krylov, M. Y. Karpeisky and R. M. Khomutov, *Bioorg. Khim.*, 1983, **9**, 838–843.
- 27 J. C. Caesar, D. V. Griffiths and J. C. Tebby, *J. Chem. Soc., Perkin Trans. 1*, 1988, 175–178.
- 28 M. Kadokura, T. Wada, C. Urashima and M. Sekine, *Tetrahedron Lett.*, 1997, **38**(48), 8359–8362.
- 29 C. E. McKenna, L. A. Khawli, W.-Y. Ahmad, P. Pham and J.-P. Bongartz, *Phosphorus Sulfur Relat. Elem.*, 1988, **37**, 1.
- 30 J. Vepsäläinen, H. Nupponen, E. Pohjala, M. Ahlgren and P. Vainiotalo, *J. Chem. Soc., Perkin Trans. 2*, 1992, 835–842.
- 31 Y. Xu, L. Qian and G. D. Prestwich, *Org. Lett.*, 2003, **5**, 2267–2270.
- 32 V. C. Nelson, *J. Labelled Compd. Radiopharm.*, 1996, **38**, 713–723.
- 33 This solution was prepared by neutralization of DMF solution of adenosine 5'-monophosphate, free acid, monohydrate with one equivalent of 55% (w/w) tetrabutylammonium hydroxide solution in water, and repeated evaporation under vacuum of the resulting solution from anhydrous DMF.
- 34 The coupling constants of this symmetrical spin system could not be measured directly from the NMR spectrum, and were determined by fitting of an AA'XX' system to the experimental data (see ESI†) using the WinDNMR program (Hans J. Reich, Department of Chemistry, University of Wisconsin).
- 35 F. Eckstein and M. Goumet, *Nucleic Acid Chem.*, 1978, **2**, 861–864.
- 36 R. Dixon and G. Lowe, *J. Biol. Chem.*, 1989, **264**, 2069–2074.
- 37 G. M. Blackburn, G. E. Taylor, G. R. J. Thatcher, M. Prescott and A. G. McLennan, *Nucleic Acids Res.*, 1987, **15**, 6991–7004.